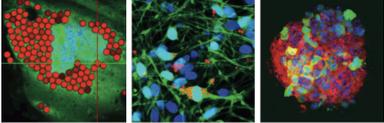


Principles of Stem Cell Biology



A one-day lecture course on what stem cells are, how they behave, how they are regulated, and how they can be used clinically.




"Principles of Stem Cell Biology" is offered by the Norwegian Center for Stem Cell Research (www.stemcellnorway.org) and the Cancer Stem Cell Innovation Center (<http://cscir-research.no/>).

MF9410 Principles of Stem Cell Biology - 1.12.2010

Course organizer: Joel C. Glover - Norwegian Center for Stem Cell Research, UiO

Course program:

0830 – 0930	Basics of stem cell biology Joel C. Glover, Norwegian Center for Stem Cell Research, UiO/OUS
0930 – 1030	Current clinical applications of stem cells in Norway Jan E. Brinchmann, Norwegian Center for Stem Cell Research, OUS/UiO
1030 – 1130	Stem cell epigenetics Philippe Collas, Norwegian Center for Stem Cell Research, UiO/OUS
1130 – 1200	Break/Lunch
1200 – 1300	MicroRNAs and stem cell regulation Jan Oxholm Gordeladze, Norwegian Center for Stem Cell Research, UiO
1300 – 1400	Tumor stem cells Stefan Krauss CAST, Norwegian Center for Stem Cell Research, OUS
1400 – 1415	Break

Current stem cell research:

1415 – 1515	Project presentations (Munthe, Moe, Andersen, Moskaug)
1515 - 1530	Break
1530 – 1615	Project presentations (Larsen, Skotheim, Glover)

STEM CELLS - BASIC CONCEPTS

Joel C. Glover
 Norwegian Center for Stem Cell Research
 CAST
 Laboratory of Neural Development and Optical Recording (NDEVOR)
 Institute of Basic Medical Sciences
 University of Oslo
joel.glover@medisin.uio.no

<http://stemcells.nih.gov/info/basics/>
<http://www.stemcellresearchfoundation.org>
<http://www.stemcell.no>

WHAT IS A STEM CELL?

A cell that can undergo self-renewing (expanding) proliferation and give rise to specialized differentiated cells

3 CONCEPTUAL CATEGORIES

Embryonic

Somatic

Tumor

3 CONCEPTUAL CATEGORIES

Embryonic
 Found in blastocyst stage embryos, can generate all tissues of the body

Somatic

Tumor

3 CONCEPTUAL CATEGORIES

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Somatic
Found in fully-formed organs, can generate multiple cell types characteristic of organ of origin.

Tumor

3 CONCEPTUAL CATEGORIES

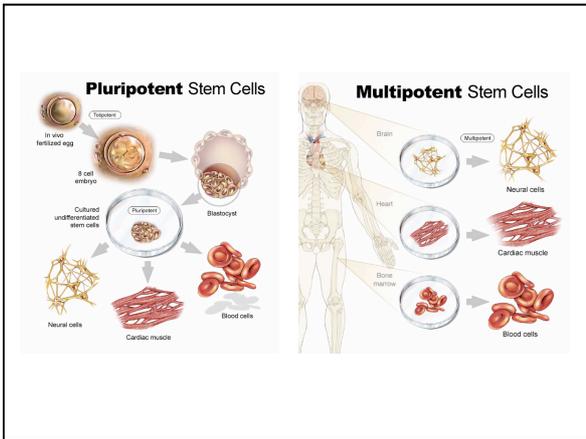
Embryonic
Found in blastocyst stage embryos, can generate all tissues of the body

Somatic
Found in fully-formed organs, can generate multiple cell types characteristic of organ of origin.

Tumor
Found in tumors, can reconstitute new tumors of same type, presumed source of metastases

THE CONCEPT OF STEM CELL POTENCY

Totipotent (entire body)	fertilized egg first few blastomeres
Pluripotent (most - all cell types)	embryonic stem cells embryonic germ cells embryonal carcinoma cells
Multipotent (several cell types)	somatic stem cells



3 CONCEPTUAL CATEGORIES

➔ **Embryonic**
Found in blastocyst stage embryos, can generate all tissues of the body

Somatic
Found in fully-formed organs, can generate multiple cell types characteristic of organ of origin.

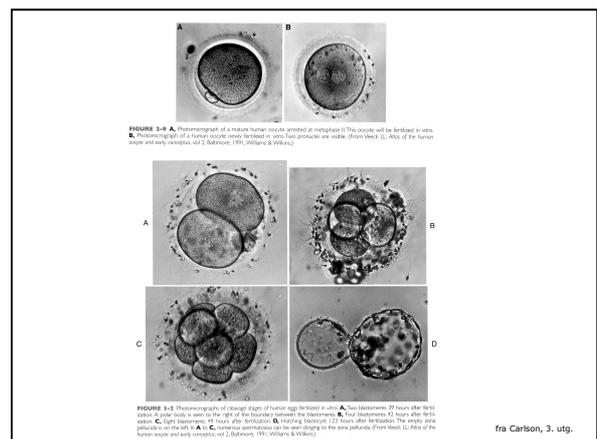
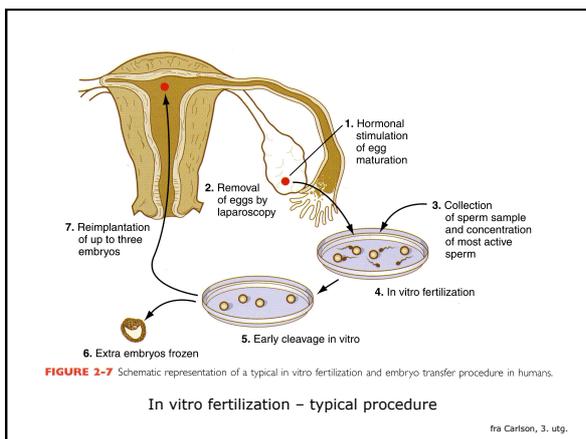
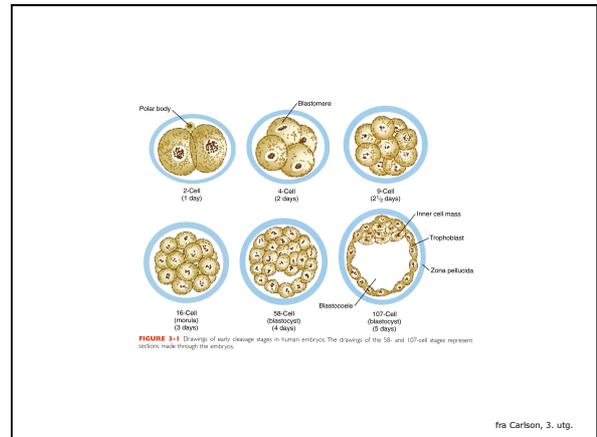
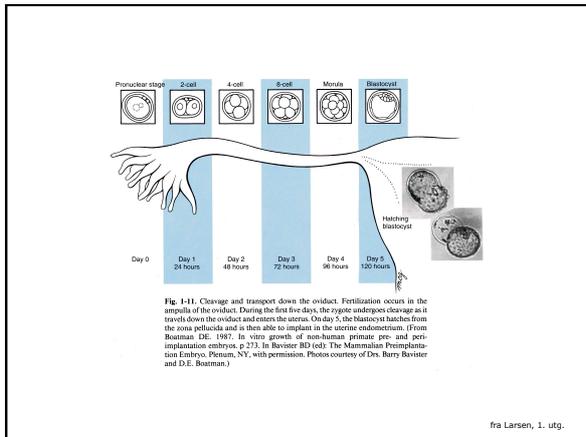
Tumor
Found in tumors, can reconstitute new tumors of same type, presumed source of metastases

HISTORICAL PERSPECTIVE

Fertilized egg + first few blastomeres are totipotent
Separated blastomere experiments of Driesch 1892

Embryonic stem cells first isolated from mouse blastocysts by Martin and Evans & Kaufman 1981
"inner cell mass"
established as expandable cell lines, are pluripotent
allowed for the generation of transgenic mice

Embryonic stem cells first isolated from human blastocysts by Thomson et al, Gearhart et al 1998
Established as expandable cell lines (first USA, now many countries including Sweden)
Requires use of human blastocysts, obtained in connection with *in vitro* fertilization for couples with fertility problems



Embryonic Stem Cell Lines Derived from Human Blastocysts

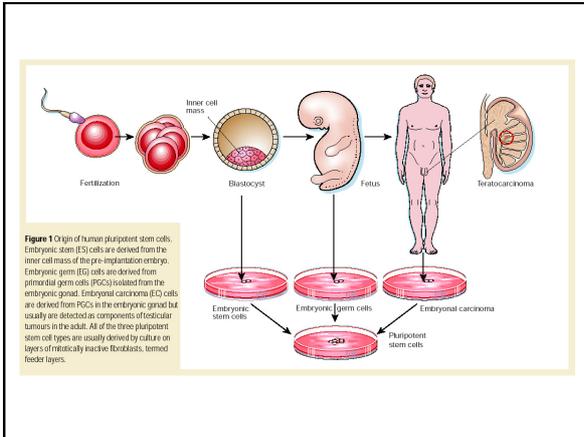
**James A. Thomson,* Joseph Itskovitz-Eldor, Sander S. Shapiro,
Michelle A. Waknitz, Jennifer J. Swiergiel, Vivienne S. Marshall,
Jeffrey M. Jones**

Human blastocyst-derived, pluripotent cell lines are described that have normal karyotypes, express high levels of telomerase activity, and express cell surface markers that characterize primate embryonic stem cells but do not characterize other early lineages. After undifferentiated proliferation in vitro for 4 to 5 months, these cells still maintained the developmental potential to form trophoblast and derivatives of all three embryonic germ layers, including gut epithelium (endoderm); cartilage, bone, smooth muscle, and striated muscle (mesoderm); and neural epithelium, embryonic ganglia, and stratified squamous epithelium (ectoderm). These cell lines should be useful in human developmental biology, drug discovery, and transplantation medicine.

www.sciencemag.org SCIENCE VOL 282 6 NOVEMBER 1998

THE CONCEPT OF STEM CELL POTENCY

<p>Totipotent (entire body)</p> <p>Pluripotent (most - all cell types)</p> <p>Multipotent (several cell types)</p>	<p>fertilized egg</p> <p>first few blastomeres</p> <p>embryonic stem cells</p> <p>embryonic germ cells</p> <p>embryonal carcinoma cells</p> <p>somatic stem cells</p>
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Embryonic stem cells: example of a potential use

Embryonic stem cells develop into functional dopaminergic neurons after transplantation in a Parkinson rat model

Lars M. Björklund^{1,2*}, Rosario Sánchez-Pernaute^{1,3}, Sangmi Chung¹, Therese Andersson^{1,4}, Iris Yin Ching Chen¹, Kevin S. P. McNaughton¹, Anna-Liisa Brownell^{1,4}, Bruce G. Jenkins¹, Claes Wahlestedt¹, Kwang-Soo Kim^{1,5}, and Ole Isacson^{1,11,12*}

¹Udall Parkinson's Disease Research Center of Excellence, ²Neuroregeneration Laboratories, and ³Molecular Neurobiology Laboratory, McLean Hospital/Harvard Medical School, 715 Mill Street, Belmont, MA 02478; ⁴Departments of ⁵Neurology and ⁶Neurobiology, Massachusetts General Hospital and Program in Neuroscience, Harvard Medical School, Boston, MA 02115; and ⁷Karolinska Institute, SE-17177 Stockholm, Sweden

Edited by Gerald D. Fischbach, Columbia University College of Physicians and Surgeons, New York, NY, and ¹⁰Department of Neurobiology, Harvard Medical School, Boston, MA 02115

2344-2349 | PNAS | February 19, 2002 | vol. 99 | no. 4

Efficient production of mesencephalic dopamine neurons by Lmx1a expression in embryonic stem cells

Stina Friling¹, Elisabet Andersson¹, Lachlan H. Thompson¹, Marie E. Jönsson¹, Josephine B. Hebsgaard¹, Evanthia Namas¹, Zhanina Aleksenko¹, Ulrika Marklund¹, Susanna Kjellander¹, Nikolaos Violakakis¹, Oriti Hovatta¹, Abdeljabbar El Manira¹, Anders Björklund¹, Thomas Perlmann^{1,2,3}, and Johan Ericson^{1,2}

¹The Ludwig Institute for Cancer Research and ²Departments of Cell and Molecular Biology, ³Neuroscience, and ⁴BioScience and Nutrition, Karolinska Institutet, 141 77 Stockholm, Sweden; and ⁵Wallenberg Neuroscience Center, Lund University, 221 84 Lund, Sweden

Contributed by Thomas M. Jessell, Columbia University College of Physicians and Surgeons, New York, NY, March 13, 2009

(received for review December 10, 2008)

PNAS | May 5, 2009 | vol. 106 | no. 18 | 7613-7618

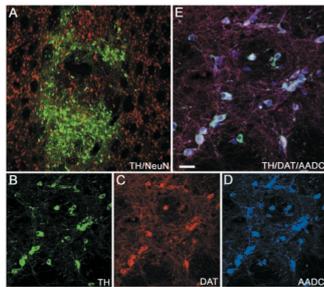


Fig. 1. Immunohistochemical staining of a graft 16 weeks after implantation of a low concentration (1,000–2,000 cells per μl) of D3 ES cells into adult 6-OHDA lesioned striatum. Numerous TH-positive neurons were found within the graft (A and B, green). All TH-positive profiles coexpressed the neuronal marker NeuN (A, red). TH (B) also was coexpressed with DAT (C, red) and AADC (D, blue), demonstrated by white triple labeling (E). (Scale bars: A, 150 μm; B–D, 50 μm; E, 25 μm.)

Björklund et al (2002) PNAS 99:2344-2349

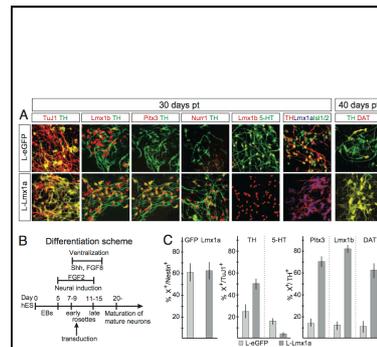


Fig. 5. Lmx1a promotes mesDA neurons in differentiating hESCs. hESC-derived neuroepithelial progenitors were infected with lentiviral (LV) vectors carrying Lmx1a or GFP and analyzed at day 30 to 40 pt. (A) In Lmx1a-infected cultures, >50% of Tu11⁺ neurons co-expressed TH at d 30 pt, compared with 25% in L-GFP-infected cultures. Most TH⁺ neurons co-expressed mesDA markers, e.g., Lmx1b, Nurr1, and DAT, whereas markers for S-HT neurons were suppressed. Few TH⁺ neurons derived from L-GFP-infected cells co-expressed mesDA markers. (B) Differentiation scheme. (C) Quantification of marker expression. Error bars indicate SD, n = 4.

Friling et al (2009) PNAS 106:7613-7618

Embryonic stem cells: example of a potential use

4694 • The Journal of Neuroscience, May 11, 2005 • 25(19):4694–4705

Development/Plasticity/Repair

Human Embryonic Stem Cell-Derived Oligodendrocyte Progenitor Cell Transplants Remyelinate and Restore Locomotion after Spinal Cord Injury

Hans S. Keirstead,¹ Gabriel Nistor,¹ Giovanna Bernal,¹ Minodora Tottoiu,¹ Frank Cloutier,¹ Kelly Sharp,¹ and Oswald Steward^{1,2,3}

¹Departments of ²Anatomy and Neurobiology, ³Neurobiology and Behavior, and ⁴Neurosurgery, Reeve-Irvine Research Center, College of Medicine, University of California at Irvine, Irvine, California 92697-4292

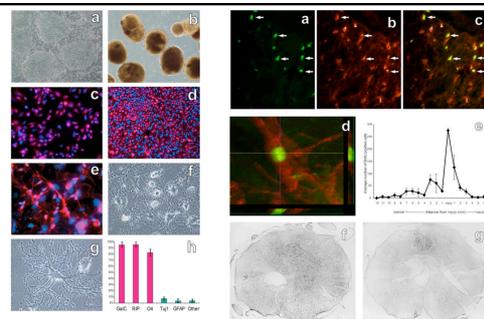
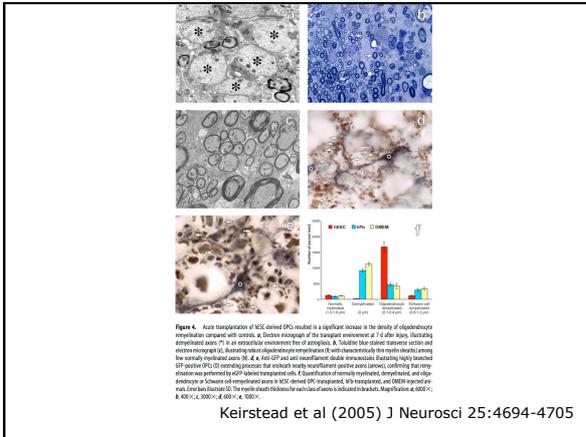


Fig. 1. Contribution of hESC to oligodendrocyte progenitor cells. Differentiated hESC neuroepithelial progenitors were infected with lentiviral (LV) vectors carrying Lmx1a or GFP and analyzed at day 30 to 40 pt. (A) In Lmx1a-infected cultures, >50% of Tu11⁺ neurons co-expressed TH at d 30 pt, compared with 25% in L-GFP-infected cultures. Most TH⁺ neurons co-expressed mesDA markers, e.g., Lmx1b, Nurr1, and DAT, whereas markers for S-HT neurons were suppressed. Few TH⁺ neurons derived from L-GFP-infected cells co-expressed mesDA markers. (B) Differentiation scheme. (C) Quantification of marker expression. Error bars indicate SD, n = 4.

Fig. 2. In vivo remyelination of hESC-derived OPCs resulted in well-oriented, dense myelination from the site of lesionation, and differentiation to mature oligodendrocytes. A, Adult human and murine OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. B, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. C, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. D, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. E, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. F, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. G, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. H, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. I, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. J, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. K, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. L, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. M, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. N, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. O, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. P, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. Q, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. R, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. S, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. T, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. U, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. V, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. W, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. X, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. Y, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord. Z, GFP⁺ OPCs (arrow) double-labeled with the myelin oligodendrocyte glycoprotein (MOG) (green) and GFP (red) (arrowhead) in the spinal cord.

Keirstead et al (2005) J Neurosci 25:4694-4705



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Somatic
Found in fully-formed organs, can generate multiple cell types characteristic of organ of origin.

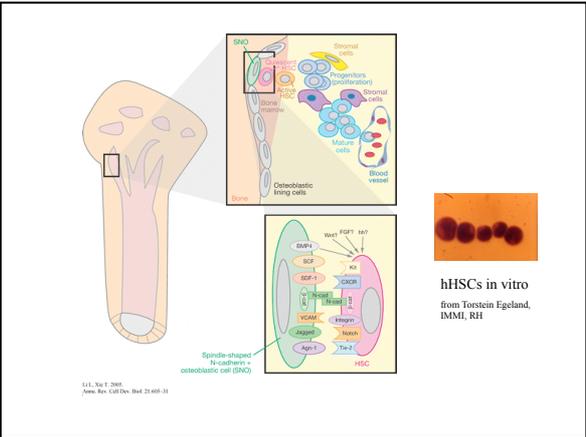
Tumor
Found in tumors, can reconstitute new tumors of same type, presumed source of metastases

HISTORICAL PERSPECTIVE

Previously known to exist in organs with obvious self-renewal (bone marrow, skin, intestinal epithelium), and in organs with some capacity to regenerate after cell loss (liver, muscle)

Previously believed NOT to exist in organs with no obvious self-renewal (like brain)

More recently demonstrated in precisely such organs (like brain)



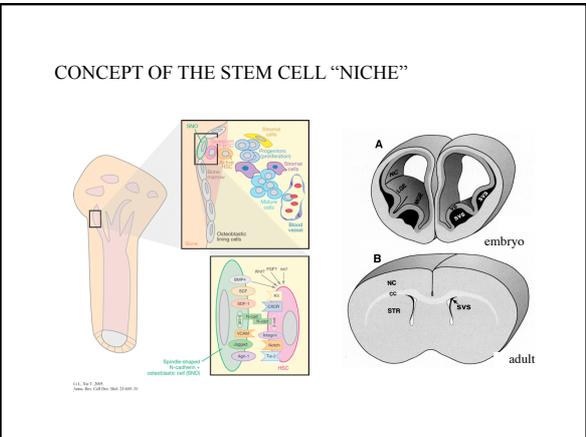
Johansson CB, Svensson M, Wallstedt L, Janson AM, Frisen J. Neural stem cells in the adult human brain. *Exp Cell Res* 1999; 253:733-736.

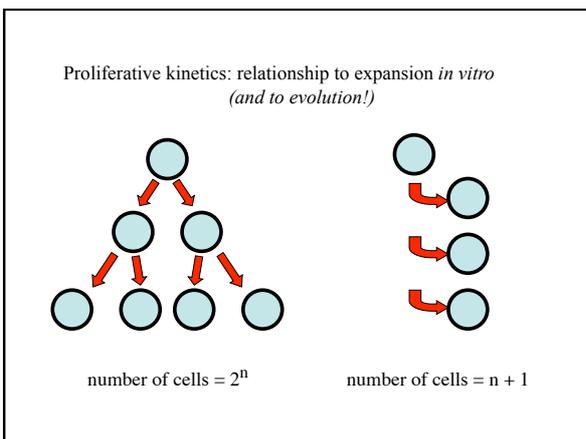
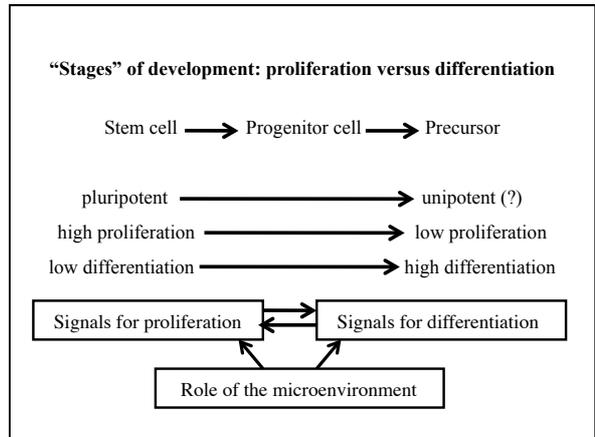
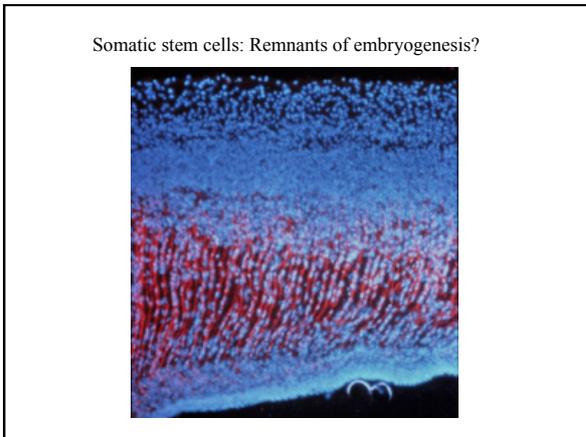
NATURE | VOL 412 | 16 AUGUST 2001 | www.nature.com

Purification of a pluripotent neural stem cell from the adult mouse brain

Rodney L. Rietze*, Helen Valcanis†, Gordon F. Brooker*, Tim Thomas*, Anne K. Voss* & Perry F. Bartlett*

*The Walter and Eliza Hall Institute of Medical Research, Royal Parade, Parkville, Victoria 3050, Australia
†Howard Florey Institute, University of Melbourne, Parkville, Victoria 3010, Australia





AN IMPORTANT QUESTION REGARDING SOMATIC STEM CELLS

What is the differentiation potential of somatic stem cells?

Organ-restricted (multipotent), or broader (pluripotent)?

Much circumstantial evidence. Requirement for definitive studies proving full differentiation to specific cell types *in vivo*.

Somatic stem cells: examples of specific uses

Hematopoietic stem cells have been used for years in the treatment of bone marrow and blood disorders such as leukemia, aplastic Anemia

Skin transplants are de facto stem cell treatments

More recent advances in regenerative medicine: Liver, connective tissue, etc.

(homotypic, as for bone marrow transplants)

In the future: Tissues derived from heterotypic stem cell sources? (for example, nerve cells from hematopoietic stem cells or from fat stem cells)

Adult human hematopoietic stem cells produce neurons efficiently in the regenerating chicken embryo spinal cord

Olafur E. Sigurjonsson*, Marie-Claude Perreault†, Torstein Egeland*, and Joel C. Glover††

*Institute of Immunology, Rikshospitalet University Hospital and University of Oslo Rikshospitalet, 0027 Oslo, Norway; and †Department of Physiology, Institute of Basic Medical Science, University of Oslo, 0319 Oslo, Norway.

Communicated by Joshua R. Sanes, Harvard University, Cambridge, MA, February 7, 2005 (received for review August 31, 2004)

Somatic stem cells: examples of specific uses

Make pluripotent stem cells!

Induced pluripotent stem cells (iPS cells): Pluripotent stem cells derived from somatic cells that have been reprogrammed to revert to a pluripotent state as in embryonic stem cells

Induction of Pluripotent Stem Cells from Mouse Embryonic and Adult Fibroblast Cultures by Defined Factors

Kazutoshi Takahashi¹ and Shinya Yamanaka^{1,2*}
¹Department of Stem Cell Biology, Institute for Frontier Medical Sciences, Kyoto University, Kyoto 606-8507, Japan
²CREST, Japan Science and Technology Agency, Kawaguchi 332-0012, Japan
 *Contact: yamanaka@fms.kyoto-u.ac.jp
 DOI 10.1016/j.cell.2006.07.024

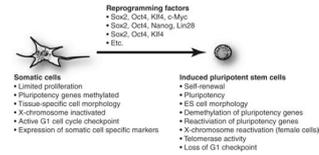


Figure 1 Reprogramming of somatic cells to induced pluripotent stem (iPS) cells. Examples of reprogramming factors are provided along with the characteristics of a typical starting somatic cell and those of an iPS cell

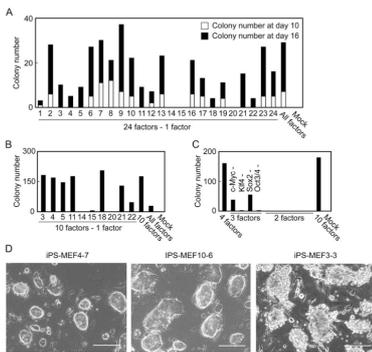


Figure 2. Narrowing down the Candidate Factors

Takahashi & Yamanaka (2006) Cell 126:663-676

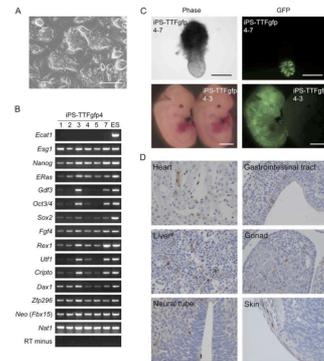


Figure 6. Characterization of iPS Cells Derived from Adult Mouse Tail-Tip Fibroblasts

Takahashi & Yamanaka (2006) Cell 126:663-676

Nature 448, 318-324 (19 July 2007) | doi:10.1038/nature05944; Reprint 22 May 2007; Published online 6 June 2007

In vitro reprogramming of fibroblasts into a pluripotent ES-cell-like state

Manica Wernig^{1,6}, Alexander Meisner^{1,6}, Ruth Foreman^{1,2,5}, Tobias Brambrink^{1,3}, Manching Ku^{1,6}, Konrad Hochdinger^{1,3}, Bradley E. Bernstein^{1,3,5} & Rudolf Jaenisch^{1,4}

Nature 454, 646-650 (31 July 2008) | doi:10.1038/nature07061; Reprint 22 May 2008; Published online 29 June 2008

Pluripotent stem cells induced from adult neural stem cells by reprogramming with two factors

Jeong Beom Kim^{1,2}, Holm Zaehres^{1,2}, Guangming Wu¹, Luca Gentile¹, Kiyomasa Koi¹, Vittorio Sebastiano¹, Marcos J. Araujo-Bravos¹, David Rhee^{1,2}, Dong Wook Han¹, Martin Zenke¹ & Hans R. Schöler¹

Neurons derived from reprogrammed fibroblasts functionally integrate into the fetal brain and improve symptoms of rats with Parkinson's disease

Manica Wernig¹, Jian-Ping Zhao¹, Jan Prusaak¹, Eva Hellmann¹, Dongsong Fu¹, Frank Soldner¹, Vanja Broccoli¹, Martha Constantine-Paton¹, Ole Isacson¹, and Rudolf Jaenisch^{1*}

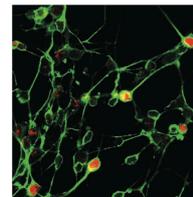
¹The Whitehead Institute for Biomedical Research, Cambridge, MA 02142, ²The McGovern Institute for Brain Research and ³Department of Biology, Massachusetts Institute of Technology, Cambridge, MA 02139, ⁴Ugall Parkinson's Disease Research Center of Excellence and Neuroregeneration Laboratories, McLean Hospital/Harvard University, Belmont, MA 02478, and ⁵San Raffaele Scientific Institute, 20132 Milan, Italy

STEM CELLS

Reprogrammed Cells Come Up Short, for Now

www.sciencemag.org SCIENCE VOL 327 5 MARCH 2010

RESPONSE TO DIFFERENTIATION SIGNALS



Work in progress. iPS cells can differentiate into functional neurons (above). But analysis of PRM2 gene expression shows they are less responsive than human ES cells to neuron-making cues (chart).

STEM CELLS

Embryonic Stem Cells Induced Pluripotent Stem Cells Exhibit Hemangioblastic Derivatives from Human Induced Pluripotent Stem Cells Exhibit Limited Expansion and Early Senescence¹⁷⁴

Guang Feng¹, Shouping Li^{1,2}, Jina Gonenkaya¹, Gustavo Gomez¹, Dohoon Kim⁴, Young Chung¹, George R. Hwang², Kwang-Doo Kim^{1,4}, Robert Lanza^{1,2,1*}

Embryonic
Advantages: Clearly pluripotent, easy to expand and differentiate, platform for many model systems for studying normal and disease mechanisms
Disadvantages: Not autologous, may cause tumors, derived from embryos

Somatic
Advantages: Autologous, already programmed towards specific cell types, lower risk of tumorigenesis
Disadvantages: Restricted potential, some are hard to get, still carry genetic disease burden

Induced pluripotent
Advantages: Autologous, greater potential, platform for in vitro disease models
Disadvantages: Harder to generate and expand, require genetic/epigenetic "harassment", may enter senescence sooner

The main message:

STEM CELL BIOLOGY STILL PRESENTS MANY CHALLENGES

What is needed is continued, integrated research into embryonic, somatic, and induced pluripotent stem cells

